Patent 260/243

## **AMENDMENTS**

IN THE CLAIMS:

Please cancel claims 1-52.

Please add the following claims:

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A method for purifying plasmid DNA, the method comprising the steps of:

- lysing cells containing the plasmid DNA, wherein said lysing step comprises:

  moving a suspension of the cells through a first passageway;

  moving a lysis solution through a second passageway;

  contacting the suspension of the cells with the lysis solution at an

  intersection, wherein the intersection is formed by the first and second

  passageway; and

  mixing the suspension of the cells and the lysis solution inside a third

  passageway to form a lysate, wherein the third passageway is
- (b) removing contaminants from the lysate, wherein the removing step comprises:

  moving the lysate through the third passageway and into a container,

  wherein the container contains a salt solution; and

  mixing the lysate in the salt solution to form a precipitate and a supernatant;

  wherein the precipitate contains the contaminants;
- (c) separating the precipitate from the supernatant; and

downstream from the intersection;

(d) recovering the plasmid DNA from the supernatant.

- The method in claim 1 wherein the third passageway comprises an in-line mixer.
- 3. The method in claim 1 wherein the container has an impeller mixer for mixing the lysate in the salt solution.
- 4. The method in claim 1 wherein the container is a holding tank having a jacket.
- 5. The method of claim 1 wherein the container has the capacity to contain a volume of about 100 to about 200 liters.
- 6. The method of claim 1 wherein the recovering of plasmid DNA yields at least about 60%.
- 7. The method of claim 1 wherein the plasmid DNA is a supercoiled plasmid DNA.
- 8. The method of claim 7 wherein the supercoiled plasmid DNA is enriched above 80%.
- 9. The method in claim 1 further comprising a cell resuspension step before the lysing step, wherein the resuspension step comprises:

recirculating the cells and a resuspension solution through a fourth passageway, wherein the cells and the resuspension solution are mixed inside the fourth passageway.

- 10. The method in claim 9 wherein the fourth passageway comprises an in-line mixer.
- 11. The method in claim 1 wherein the step of removing the contaminants further comprises chilling the precipitate and supernatant in the container at 4 degree Celsius for a time between about 6 to about 12 hours, and wherein the chilling step is before the step of separating the precipitate from the supernatant.

- 12. The method in claim 1 wherein the suspension of cells, the lysis solution, and the lysate are moved at a flow rate of about 1 liter/min.
- 13. The method in claim 1 wherein the salt solution is a non-acidic salt solution.
- 14. The method in claim 1 wherein the salt solution is unbuffered salt solution.
- 15. The method in claim 1 wherein the salt solution has a salt concentration of greater than 5M.
- 16. The method of claim 13 wherein the non-acidic salt solution comprises a salt selected from a group consisting of acetate salt and chloride salt.
- 17. The method of claim 16 wherein the acetate salt has a concentration of about 8M.
- 18. The method of claim 17 wherein the acetate salt is a mixture of potassium acetate and ammonium acetate.
- 19. The method of claim 18 wherein potassium acetate is at a concentration of about 1M and ammonium acetate is at a concentration of about 7M.
- 20. The method of claim 13 wherein the non-acidic salt solution has a pH between 7 to 9.
- 21. The method in claim 13 wherein the non-acidic salt solution comprises a mixture of two salts, the salts being selected from a group consisting of potassium acetate, potassium chloride, sodium chloride, and ammonium acetate.
- 22. The method of claim 1 wherein the lysis solution is basic.
- 23. The method of claim 22 wherein the lysis solution has a pH of about 12-13.

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- 24. The method of claim 23 wherein the lysis solution comprises 0.2N sodium hydroxide and a detergent selected from a group consisting of SDS, triton X, Tween, sarkosyl, and NP-40.
- 25. The method in claim 1 wherein the precipitate comprises contaminants selected from the group consisting of RNA, chromosomal DNA, lipids, and protein.
- 26. The method in claim 1 wherein the step of recovering the plasmid DNA from the supernatant comprises exposing the supernatant to column chromatography.
- 27. The method in claim 26 wherein the step of exposing the supernatant to column chromatography involves the use of a hydrophobic interaction column.
- 28. The method in claim 27 wherein the hydrophobic interaction column comprises a resin selected from a group consisting of: Octyl Sepharose 4 Fast Flow, Phenyl Sepharose, Butyl Sepharose, Phenyl 650-S.
- 29. The method in claim 26 wherein the step of exposing the supernatant to column chromatography comprises:

adjusting a salt concentration of the supernatant to have at least 2M ammonium sulfate; flowing the adjusted supernatant through an hydrophobic interaction column; and eluting the plasmid DNA with an eluent solution having less than 2M ammonium sulfate.

- 30. The method in claim 29 wherein the eluent solution has ammonium sulfate at a concentration of less than or equal to 1.6M.
- 31. The method in claim 26 wherein the step of exposing the supernatant to column chromatography comprises:

adjusting a salt concentration of the supernatant to have about 1.6M to about 1.75M ammonium sulfate;

flowing the adjusted supernatant through an hydrophobic interacting column; and collecting a flow through eluate flowing from the hydrophobic interacting column.

- 32. The method in claim 27 wherein the step of exposing the supernatant to column chromatography further involves the use of an anionic exchange column.
- 33. The method in claim 32 wherein the use of an anionic exchange column comprises eluting plasmid DNA via a step gradient.
- 34. The method in claim 32 wherein the anionic exchange column comprises a resin having a particle size of 20-40 microns.
- The method in claim 32 wherein the anionic exchange column comprises a resin selected from a group consisting of: Fractogel EMD TMAE (650-S), Fractogel (R) EMD TMAE Hicap,

  Q Sepharose 4 Fast Flow, DEAE 650-S
- 36. The method in claim 32 wherein the plasmid DNA is eluted from the anionic exchange column with a salt solution selected from a group consisting of: about 1.9 M ammonium sulfate and at least 0.7M NaCl.
- 37. The method in claim 26 wherein the step of exposing the supernatant to column chromatography further comprises the steps of:

passing the supernatant through the anionic exchange column; wherein the plasmid DNA binds to the anionic exchange column;

eluting the plasmid DNA in a first eluate from the anionic exchange column;

adjusting a salt concentration of the first eluate to contain at least 2M ammonium sulfate; passing the first eluate through the hydrophobic interaction column; wherein supercoiled plasmid DNA binds to the hydrophobic interaction column; and

eluting the supercoiled plasmid DNA in a second eluate from the hydrophobic interaction column via an eluent having less than 2M ammonium sulfate.

- 38. The method of claim 37 wherein the plasmid DNA in the first eluate is eluted from the anionic exchange column with a salt solution selected from a group consisting of: about 1.9 M ammonium sulfate and at least 0.7M NaCl.
- 39. A device for purifying plasmid DNA from cells containing said plasmid DNA, the device comprising:
  - a first in-line mixer for resuspending the cells into a homogenous cell suspension;
- a second in-line mixer in fluid connection with the first in-line mixer for mixing the homogenous cell suspension with a lysis solution to form a lysate;
  - a container for holding a salt solution, wherein the holding tank comprises:
  - a mixer for mixing a salt solution in the holding tank with the lysate flowing from
    the second in-line mixer to form a precipitate and a supernatant; and
    a container outlet from which the precipitate and supernatant are removed from
    the tank; and
- at least one pump for causing the cells to flow through the first in-line mixer and the homogeneous cell suspension and lysis solution to flow through the second in-line mixer.
- 40. The device in claim 39 wherein the container has a capacity to hold a volume of about 100 to about 200 liters.

41. The device in claim 39 further comprising a chiller that chills the container to at least 10 degree Celsius or below.

The device in claim 41 wherein the container holding tank having a jacket and wherein the chiller comprises a recirculating chiller connected to the jacket.

- 42. The device in claim 39 further comprising:
- a reservoir for holding a resuspension solution and the cells, and wherein the reservoir is in fluid connection with the first in-line mixer.
- The device in claim 42 wherein the first in-line mixer comprises a tubing having:
  - a tubing inlet connected to the reservoir, said tubing inlet allowing the resuspension solution and the cells to flow into the tubing from the reservoir; and
  - a tubing outlet directed back at the reservoir such that an effluent from the tubing flows back into the reservoir.
- 44. The device in claim 39 wherein the second in-line mixer comprises:
- a first inlet whereby the homogenous cell suspension enters the second in-line mixer through the first inlet;
- a second inlet whereby the lysis solution enters the second in-line mixer through the second inlet; and

wherein the first and second inlet joins at an intersection to form an outlet.

- 45. The device of claim 44 wherein the intersection is a Y-shaped intersection.
- 46. The device in claim 44 wherein a static mixer is positioned downstream of the outlet.

The device in claim 44 wherein the outlet is connected to a tubing that coils around the container.

- 47. The device in claim 39 wherein container further comprises a filter connected to the container outlet for filtering the precipitate.
- 48. The device in claim 47 wherein the holding tank is in fluid connection with a column chromatography system.
- 49. A device for purifying plasmid DNA from cells containing said plasmid DNA; the device comprising:
  - a first in-line mixer for resuspending the cells into a homogenous cell suspension;
- a second in-line mixer in fluid connection with the first in-line mixer for mixing the homogenous cell suspension with a lysis solution to form a lysate; and
- a third in-line mixer in fluid connection with the second in-line mixer for mixing the lysate with a salt solution to form a precipitate and a supernatant.
- 50. A method of purifying plasmid DNA from a cell, the method comprising the steps of:
  lysing the cell with a lysis solution to form a lysate solution;

removing contaminants from the lysate solution by precipitating the contaminants from a supernatant of the lysate solution;

recovering plasmid DNA from the supernatant using a hydrophobic interaction column.

The method in claim 50 wherein the hydrophobic interaction column comprises a resin selected from a group consisting of: Octyl Sepharose 4 Fast Flow, Phenyl Sepharose, Butyl Sepharose, Phenyl 650-S.

52. The method in claim 50 wherein the step of recovering the plasmid DNA from the supernatant comprises the steps of:

adjusting a salt concentration of the supernatant to have at least 2M ammonium sulfate; flowing the adjusted supernatant through the hydrophobic interaction column; and eluting the plasmid DNA with an eluent solution having less than 2M ammonium sulfate.

- 53. The method in claim 52 wherein the eluent solution has ammonium sulfate at a concentration of less than or equal to 1.6M.
- 54. The method in claim 50 wherein the step of recovering plasmid DNA from the supernatant comprises the steps of:

adjusting a salt concentration of the supernatant to have about 1.6M to about 1.75M ammonium sulfate;

flowing the adjusted supernatant through the hydrophobic interacting column; and collecting a flow through cluate flowing from the hydrophobic interacting column.

- 55. The method in claim 50 wherein the step of recovering plasmid DNA from the supernatant further involves the use of an anionic exchange column.
- 56. The method in claim 55 wherein the use of an anionic exchange column comprises the step of cluting plasmid DNA via a step gradient.
- 57. The method in claim 55 wherein the anionic exchange column comprises a resin having a particle size of 20-40 microns.

- 58. The method in claim 55 wherein the anionic exchange column comprises a resin selected from a group consisting of: Fractogel EMD TMAE (650-S), Fractogel (R) EMD TMAE Hicap, Q Sepharose 4 Fast Flow, DEAE 650-S
- 59. The method in claim 55 wherein the plasmid DNA is eluted from the anionic exchange column with a salt solution selected from a group consisting of: about 1.9 M ammonium sulfate and at least 0.7M NaCl.
- 60. The method in claim 50 wherein the step of recovering plasmid DNA from the supernatant comprises the steps of:

an eluent having less than 2M ammonium sulfate.

passing the supernatant through an anionic exchange column; wherein the plasmid DNA binds to the anionic exchange column;

eluting the plasmid DNA in a first eluate from the anionic exchange column;
adjusting a salt concentration of the first eluate to contain at least 2M ammonium sulfate;
passing the first eluate through the hydrophobic interaction column; wherein supercoiled
plasmid DNA binds to the hydrophobic interaction column; and
eluting the supercoiled plasmid in a second eluate from the hydrophobic interaction column via